



25 years in 25 articles

CH₄ and **N**₂**O Emissions in a Rice Field:** First Measurements in the Uruguayan Productive System

Emisiones de CH₄ **y N**₂**O en un arrozal:** primeras medidas en el sistema productivo uruguayo

Irisarri, P.¹; Pereyra, V.²; Fernández, A.²; Terra, J.³; Tarlera, S.²

¹Universidad de la República, Facultad de Agronomía, Departamento de Biología Vegetal, Montevideo, Uruguay ²Universidad de la República, Facultad de Química, Laboratorio de Ecología Microbiana y Ambiental, Montevideo, Uruguay ³Instituto Nacional de Investigación Agropecuaria (INIA), Treinta y Tres, Uruguay

Article originally published in:

Agrociencia (Uruguay). 2012;16(2):1-10. doi: 10.31285/AGRO.17.533

Abstract

Irrigated rice fields are major sources of two important greenhouse gases (GHG), methane and nitrous oxide. As an initial step towards obtaining local information, emissions of CH₄ and N₂O from rice paddy soil were measured by the static chamber technique in greenhouse and field experiments conducted in eastern Uruguay. In the greenhouse experiment, the effect of two flooding moments (21 and 45 days after emergence) and nitrogen fertilization (0 and 50 kg N ha⁻¹) on gas emissions was studied. Early flooding and nitrogen fertilization tended to increase N₂O emissions. In the field experiment, effect of winter soil cover crop and nitrogen fertilization (0 and 82 kg N ha⁻¹) were tested. Higher CH₄ fluxes were observed mainly during the reproductive stage of the plant in the N-fertilized treatment with ryegrass winter crop. N₂O flux peaked at flushing. Results indicate that the use of cover crops might increase GHG emissions during the rice cycle. Despite differences in agronomic management practices employed in Uruguay, CH₄ and N₂O fluxes are within magnitudes previously reported for rice fields worldwide.

Keywords: rice paddy soil, greenhouse gases, N fertilization

Resumen

Los arrozales son fuente de dos importantes gases de efecto invernadero (GEI), metano y óxido nitroso. Como un paso inicial hacia la obtención de información local, se midieron las emisiones de CH₄ y N₂O del suelo y de las plantas de arroz mediante la técnica de la cámara estática en experimentos en invernáculo y a campo en el este de Uruguay. En el experimento en invernáculo, se estudió el efecto del momento de inundación (21 y 45 días después de la emergencia) y de la fertilización nitrogenada (0 y 50 kg N ha⁻¹) sobre las emisiones. La inundación temprana y la fertilización nitrogenada tendieron a aumentar las emisiones de N₂O. En el experimento a campo, se estudió el efecto de la cobertura invernal y de la fertilización nitrogenada (0 y 82 kg N ha⁻¹). Se detectaron mayores flujos de CH₄ durante la etapa reproductiva de la planta en el tratamiento fertilizado con cobertura invernal previa de raigrás. El flujo de N₂O fue máximo después de los baños. Los resultados indican que el uso del cultivo de cobertura podría incrementar las emisiones de GEI durante el ciclo del arroz. A pesar de las distintas prácticas de manejo del cultivo empleadas en Uruguay, los flujos de CH₄ y N₂O se encuentran dentro de los valores informados previamente para arrozales de otras partes del mundo.

Palabras clave: suelo inundado cultivado con arroz, gases de efecto invernadero, fertilización N



Irisarri P, Pereyra V, Fernández A, Terra J, Tarlera S. CH₄ and N₂O Emissions in a Rice Field: First Measurements in the Uruguayan Productive System. Agrociencia Uruguay [Internet]. 2022 [cited dd mmm yyyy];26(NE2):e1083. doi:10.31285/AGRO.26.1083

☐ Correspondence

Silvana Tarlera, starlera @fq.edu.uy



Introduction

Agricultural soils are important global sources of methane (CH₄) and nitrous oxide (N₂O) (IPCC, 2007). In Uruguay, the CO₂ captured by forests almost doubles their emission, which is why CH₄ and N₂O are the main greenhouse gases (GHGs). It is estimated that agriculture is responsible for 92.6% of the emissions of CH₄ and almost all of those of N₂O (MVOTMA and others, 2010). Greenhouse gases (GHGs) have different heating capacities, based on their impact on radiant energy and their duration in the atmosphere compared to the reference gas, carbon dioxide (CO₂). CH4 and N₂O have a 25-fold and 298-fold higher heating potential than CO₂ respectively, for a 100-year time scale (IPCC, 2007).

Rice is the main irrigated crop in Uruguay, 55% of the cultivated area is in the east of the country and approximately 90% of production is exported (ACA, 2011). The national inventory of emitted GHGs estimates that rice cultivation is responsible for 4% of the CH₄ emitted in Uruguay (MVOTMA and others, 2010). One of the environmental challenges of systems that include flood rice cultivation is to reduce the emission of GHGs produced mainly by microbial activities. Emissions are strongly influenced by nitrogen fertilization, the management of soil and crop residues during fallowing and the management of irrigation water. Rice cultivation is considered the main global anthropogenic source of CH₄ (Jacobson, 2005). Emissions of CH₄ depend on rice cultivars (Kerdchoechuen, 2005), but are also increased by the incorporation of organic matter into the soil (Yagi and Minami, 1990; Bronson and others, 1997), and their mitigation is based on reducing the time the crop remains flooded (Yagi and others, 1996; Cai and others, 1997).

N₂O occurs mostly at the interface between dry and flooded soil (Cai and others, 2001; Xing and others, 2002). Its emission depends on soil drainage (Towprayoon and others, 2005) and is stimulated by nitrogen fertilization (Bronson and others, 1997; Crutzen and Lelieveld, 2001). The emission of CH₄ results from the balance between the activities of methanogenic archaea, strictly anaerobic, and methanotrophic, aerobic bacteria (Macalady and others, 2002). On the other hand, N₂O is the product of incomplete microbial transformations of nitrogen compounds incorporated into the soil as a fertilizer, in oxic (nitrification) or anoxic conditions (mainly denitrification and denitrifying nitrification) (Smith and others, 2003; Baggs and Philippot, 2011). Rice cultivation in Uruguay is unique in the world as it shares the use of soils with pasture for livestock and other crops in rotation (ACA, 2011). Rice integrated into these systems produces high yields with the application of low doses of agrochemicals and preserves soil guality (Deambrosi, 2003; Méndez and others, 2003). Global trade presents increasing demands on the environmental impacts of production processes and their documentation, including the requirement of water use, environmental destination of agrochemicals, and GHG emissions among others (Itoh and others, 2011). This study aimed to obtain the first local emission data of CH4 and N₂O in our country's particular rice production system. In addition, a first approximation was made of the impact of some management practices, nitrogen fertilization, water management, and winter covers, on the fluxes of these GHGs.

Material and methods

Greenhouse experiment

The greenhouse experiment was carried out at the National Institute of Agricultural Research (INIA) of the department of Treinta y Tres. Rice (*Oryza sativa* L., cultivar "El Paso 144") was planted in plastic crates with soil taken from the upper 0.30 m in the Experimental Unit "Paso de la Laguna" of INIA Treinta y Tres, with the following characteristics: silty loam texture, pH (H₂O) 5.2; N-NH₄+ 0.3 mmol L⁻¹; organic C 30-35 g kg⁻¹ and organic matter 50-55 g kg⁻¹. The apparent density of the soil was 1.36 g cm⁻³. This experiment was carried out to assess the effects of the moment of flooding and nitrogen fertilization on emissions. A random plot design was used, with four replications and two chambers in each crate.

Each crate was filled with 52 kg of soil and basal fertilization of 120 kg ha⁻¹ of ammonium phosphate (18-46-0) was applied. Rice was seeded at a density of 180 kg ha⁻¹ and seedlings were irrigated weekly to field capacity. Nitrogen treatment consisted of the application of urea 50 kg ha⁻¹ to the seedlings and at 21 days after the emergence (DAE). Rice plants emerged nine days after sowing. The water management treatment consisted of two dates of flood establishment, 21 DAE (early flood) and 45 DAE (late flood). The water level during the flood was kept at 5-6 cm above the ground until harvest, which was performed at 134 DAE.

Field experiment

The experiment was carried out at INIA's Paso de la Laguna Experimental Unit (33°16′S, 54°16′W)

during the 2008-09 rice harvest to learn about the effect of the inclusion of winter cover crops on CH_4 and N_2O emissions during rice cultivation. The soil was Albic Natraqualf (USDA, 1998) with three previous years of rest without rice. The physicochemical characteristics of the soil are shown in Table 1.

Table 1. Soil properties in the field experiment.

$\mathrm{pH}\left(\mathrm{H_{2}O}\right)$	Organic C	Total N	P Bray	Available K
	(g kg ⁻¹)	(g kg ⁻¹)	(µgg ⁻¹)	(m eq100g ⁻¹)
5.4±0.17	19.0±1.7	1.7±0.1	7.8±0.9	14.6±4.8

The treatments were the factorial arrangement of two soil managements during the winter, ryegrass (Lolium multiflorum Lam.) as cover or bare soil, and two doses of nitrogen fertilization, 0 and 82 kg N ha-1. The design was of random plots of 10 m x 9.2 m with four replications and two chambers in each plot. The ryegrass was sown on March 30 at a density of 20 kg of seeds ha-1, 10 days after an application of 1.5 kg ia ha⁻¹ of glyphosate (Terra and others, 2009). The bare soil treatment (without vegetation) received a second dose of glyphosate on June 20, apart from that of March 20. In both treatments, the chemical fallow began on September 19 with an application of 2.5 kg ha⁻¹ of glyphosate. The total dry matter harvested from ryegrass was 4940 kg ha-1 with a C/N ratio of 47/1 (Terra and others, 2009).

The soil tillage was carried out the previous summer (January 2008) and consisted of a heavy eccentric run, two runs of disc track, and two runs of *landplane*.

The cultivation of rice (cv. INIA Olimar) was installed on October 13 with no-till at a density of 150 kg of seed ha⁻¹. The flooding was performed 22 DAE and a 10 cm sheet of water was maintained until five days before harvest. Two pre-flood flushings of the crop were performed at 1 and 4 DAE.

The fractional nitrogen fertilization consisted of the application, at sowing, of ammonium phosphate (22 kg N ha⁻¹, 23 DAE), urea to the tillering (21 DAE) 30 kg N ha⁻¹ on dry soil, and to the primordium (51 DAE) 30 kg N ha⁻¹. Treatments without N did not receive any application of nitrogen fertilizer.

Yield parameters such as 13% adjusted grain weight, number of stems m⁻², grains per panicle, dry matter in flowering stages, and primordium, were evaluated according to Terra and others (2009). The estimation of the chlorophyll content of the rice leaves was measured in the most developed upper

leaf with a SPAD 502 Plus Chlorophyll meter (Terra and others, 2009).

The weather information was recorded at the station located at "Paso de la Laguna".

The average temperature was 21.5 °C during the crop cycle (Figure 1) and the precipitation 576 mm, of which only 36 mm were recorded before flooding. The thermal amplitude had an average value of 12 °C throughout the rice cycle.

Figure 1. Daily temperature during the rice production cycle. Black rhombuses indicate the average temperature, squares the maximum and triangles



Sampling and flux measurements of CH4 and N_2O

The gas fluxes emitted were monitored using the static closed chamber technique described for rice by Lindau and others (1991) on the dates indicated for each experiment in Figures 3 and 4 and between 13 and 15 h. The chambers consisted of stainless steel bases of 40 cm in diameter and 20 cm in height partially inserted (5 cm) in the soil that remained installed throughout the cultivation cycle. On each sampling date, 60 cm high acrylic cylinders were placed on the bases with a water seal to prevent the escape of gases. The chambers had a battery-operated fan that was switched on five minutes before each measurement to ensure the homogeneity of the atmosphere inside the chamber and a device to balance the internal and external pressure (Figure 2). Gas samples from inside the chambers were taken with 25 mL plastic syringes at 0, 30, and 60 minutes and stored in vacuum tubes (10mL) until analysis. The temperature of the chambers, the depth of flood water, and the height of air space in each chamber were recorded to calculate the gas fluxes over time. Concentrations of CH₄ were anawith Chrompack CP 9001 lyzed а gas



chromatograph equipped with an FID detector (flame ionization detector). The analysis of N2O was performed with a modified 14B Shimadzu gas chromatograph with an ECD detector (electronic capture) described in Perdomo and others (2009). The emission rate of both gases was calculated according to Watanabe and others (2000): $F = \tilde{n}.h(dC/dt)$; where F corresponds to the emission rate of N-N₂O or C-CH₄ in g ha⁻¹ d⁻¹; \tilde{n} is the density of N-N₂O or C-CH₄ corrected by the temperature inside the sampling chamber; h is the height of the chamber from the ground or the water level, and dC/dt is the increase in the concentration of N₂O or CH₄ inside the chamber over time. Before calculating the emission rates, the existence of a linear relationship between the concentration of the corresponding gas and time was confirmed for each case. The emission rate obtained for the replications of each treatment was averaged to determine the final emission value per treatment.

Figure 2. Photo of acrylic cylinders used for gas measurements.



The emitted seasonally integrated flux (Esif) was calculated from the areas under the gas emission figures for the entire time of rice cultivation for each of the chambers.

Soil Analysis

Random composite samples of eight cylinders 0-10 cm deep were collected to determine nitrate (NO₃⁻). Samples were dried in a forced-air oven at 40°C, passed through 2 mm sieve and NO₃⁻ was analyzed by colorimetry after extraction with 2M KCl at 5:1

ratio. The content of NO₃⁻ was determined after reduction through a Cd column (Griess-Ilosvay reaction; Mulvaney, 1996).

Figure 3. Evolution of CH₄ and N₂O flux and content of NO₃⁻ in the soil in the greenhouse experiment according to the moment of flooding (DAE: days after emergence) and nitrogen fertilization. (a) Emission of CH₄; (b) emission of N₂O; (c) concentration of NO₃⁻ in soil. Treatments: 21 DAE (empty circles); 21 DAE + N (full circles); 45 DAE + N (full squares). Arrows indicate the application time of the fertilizer. Vertical lines indicate the moment of flooding: (—) 21 days and 45 days. Each point of the graphs corresponds to the mean of the flux calculated from eight static chambers (two in each





Statistical analysis

The emission data obtained in the greenhouse and field experiments were evaluated by adjusting Mixed Effects Models using Software R (2009). For the analysis of the results obtained in greenhouse, treatments (combinations of different moments of



flooding and levels of nitrogen fertilization), the time covariate, and their interaction were considered as fixed effects, while replications were considered as random effects. Field experiment data were analyzed considering the time covariate, winter covers, the level of nitrogen fertilization, and the interaction between the two last variables as fixed effects.

The adjustment of alternative models to the data groups was compared using variance analysis (ANOVA) and the most appropriate model was selected. A variance analysis (P=0.05) was applied to the results obtained with the final adjusted model.

Results and discussion

Greenhouse experiment

Figure 3a shows the CH₄ fluxes from the greenhouse experiments. No emissions of CH₄ were detected during the rice vegetative growth period (0 to 50 DAE), regardless of the flooding date. This period covered up to 34 days after flooding in early flood treatment and five days in late flooding. The rice was at advanced tillering at 50 DAE in both treatments when the emission of CH₄ was detected.

For early flood treatment, the first emission value was detected five weeks after the flood, while for late flood it was two weeks after the crop was flooded. On that date (64 DAE), with both treatments in the flowering initiation stage, there were no significant differences between the emissions. At 104 DAE, rice was in the flowering stage in the late flood treatment, but plants of the early flooding were more advanced, in the ripening stage. The practice of advancing the flooding has been reported as promoting crop maturity (Deambrosi, 2003). Coinciding with our results, it has been reported that about 90% of the total CH₄ in the entire crop cycle is emitted in the flowering, due to the maximum increase in biomass at that stage (Holzapfel-Pschorn and others, 1986; Schütz and others, 1989; Neue and others, 1997).

Table 2 shows the positive effect of nitrogen fertilization on the rice yield with early flooding. However, differences were not significant in the CH₄ fluxes between these treatments. The reported results on the effect of mineral N-fertilizers and the emission of CH₄ in flooded rice fields are contradictory (Wassmann and others, 1993). Different studies revealed that it is a relatively complex effect that is not yet fully understood (Bodelier and others, 2000), either because it stimulates or represses the main microbial populations involved in the generation and oxidation of CH₄. It should also be considered that fertilization not only affects microorganisms but also plants, adding complexity to the final result. These results suggest that the CH₄ fluxes are dependent on the plant development stage and that flooding would have an indirect influence on the emission of CH₄ when regulating the crop cycle.

Figure 4. Evolution of the flux of CH₄ and N₂O and content of NO₃⁻ in the soil in the field experiment according to the previous winter cover (DAE: days after emergence) and nitrogen fertilization. (a) Emission of CH₄; (b) emission of N₂O; (c) concentration of NO₃⁻ in soil. Treatments: Ryegrass (empty circles); Ryegrass+N (full circles); Soil without vegetation (empty squares); Soil without vegetation + N (full squares). Arrows indicate the application time of the fertilizer. Vertical lines indicate the moment of flooding: (—) 21 days DAE,

flushing (-.-) and drainage (...) before harvest. Each point of the graphs corresponds to the mean of the flux calculated from eight static chambers (two in each plot).





 Table 2. Rice grain yield in the greenhouse experiment¹.

Treatment ²	kg ha ⁻¹
21 DAE	$_{6709\pm666}$ b
21 DAE+N	8084 ± 897 a
45 DAE+N	$8457\pm807~a$

¹The yield values and their error are presented. Different letters indicate significant differences (p<0.01).

²Treatment: combination of moment of the flood establishment (DAE: days after emergence) and application of nitrogen fertilizer (+ N).

The highest peak of N₂O (38 g N ha⁻¹ day⁻¹) was recorded in the early flood treatment the day after urea fertilization and flooding (Figure 3b) and this flux was significantly different from that of the other treatments (p<0.01). This event coincided with a decrease in soil NO₃-concentration (Figure 3c). This emission peak of N₂O could be attributed to the application of N-fertilizer if compared to the treatment with the same flood date but unfertilized and in which the emissions of N₂O remained low and constant throughout the crop cycle. However, the flooding itself contributed to this emission increase if we compare both fertilized treatments. On the same date, for the treatment that remained unflooded, there was a lower flux of N₂O (14 g N ha⁻¹ day⁻¹).

During the period without flooding, emissions of N₂O were probably due to the nitrification of NH₄⁺. When the soil was flooded early (21 DAE), denitrifying microorganisms acted on the pool of NO3⁻ released by nitrification producing N2O. Denitrification is normally considered the main source of N₂O in soils (Kravchenko and Yu, 2006). The flux of N₂O decreased dramatically after the soil was flooded permanently (10 cm water sheet) (Figure 3b), which can be attributed to the recapture of N₂O and reduction to N₂ under strictly anaerobic conditions. In fact, one of the currently studied ways to mitigate N2O emissions is to increase the reduction from N₂O to N₂ (Baggs and others, 2010). In the 45 DAE treatment, the flooding was done with the rice in late tillering, when the NO₃- available for denitrification was lower probably due to greater absorption by the plants. The N₂O fluxes were barely detectable during the rest of the rice crop cycle (Figure 3b).

Field experiment

The emission patterns of CH₄ were similar for rice cultivation with the two soil managements in the previous winter: ryegrass and soil without vegetation (Figure 4a). However, the records of CH₄ emissions began two weeks after flooding (34 DAE) on plots that had had ryegrass as cover, while at that time the emission was negligible for sown plots in soil without vegetation. As in the greenhouse experiment, CH₄ flux increased in the reproductive phase (onset of flowering, 78 DAE) and the maximum peak was recorded at 93 DAE (flowering) in all treatments. These results agree with previous reports that showed a positive correlation between a high production of CH₄ and the flowering stage, due to the increase of organic root exudates in this stage of the plant (Holzapel-Pschorn and others, 1986).

Several studies have emphasized that increased fluxes of CH₄ in late stages of plant growth would be caused by the proliferation of root exudates or products of root autolysis (Holzapfel-Pschorn and others, 1986; Lindau and others, 1991; Neue and Sass, 1994; Chidthaisong and Watanabe, 1997). Table 3 indirectly illustrates this point since a considerable increase in the dry matter of all treatments was observed during flowering. As a consequence of this increase in biomass, there was a greater availability of decomposable carbon from root exudates which, in turn, serve as a source of carbon and energy to microflora. After harvest (149 and 163 DAE, for bare soil and ryegrass respectively) very few emissions were recorded.

A significant interaction (p=0.01) was observed between the cover with ryegrass and the N-fertilization, with the highest fluxes of CH₄ for the treatment of fertilized rice and with ryegrass as winter cover. Different organic aggregates are generally considered to stimulate the flux of CH₄ by increasing the carbon supply for methanogens (Yagi and others, 1996; Bronson and others, 1997; Wassmann and others, 2000). Especially, if the incorporated material has a high C/N ratio, as in the case of ryegrass stubble. The plant absorption of nutrients leaves less N available to microorganisms and therefore N could be limiting bacterial activity.

Rice yield from ryegrass treatment, regardless of the N dose, was lower (Table 3).

Treatments	Grain Yield (kgha-1 ₁	Dry matter at panicle initiation (kg ha ⁻¹)	Dry matter in flowering (kg ha ⁻¹)	Chlorophyll content at panicle initiation (SPAD units)	Chlorophyll content in flowering (SPAD units)	Number of panicles m²	Grains per panicle
Without vegetation Without vegetation Ryegra Ryegras +	10370 ± 925 a 11233 ± 324 a 8777 ± 1543 b 9870 ± 904 ab	5637 ± 1781 a 6194 ± 531 a 1837 ± 262 c 3405 ± 706 b	9840 ± 1832 b 13375 ± 2839 a 5052 ± 1011 c 7199 ± 564 bc	35.8 ±1.2 bc 31.8 ± 2.4 c 36.6 ± 2.2 a	36.2 ± 2.6 a 33.2 ± 2.4 b 39.9 ± 1.7 a 40.8 ± 1.1 a	581 ± 146 a 510 ± 62 a 313 ± 59 b 352 ± 63 b	117 ± ⁸ ab 100 ± 20 b 123 ± 16 a 133 ± 8 a

Both treatments showed less accumulation of dry matter, number of panicles per m² and grains per panicle. In contrast, the tendency of chlorophyll content was opposite, with the highest values observed in ryegrass treatments regardless of fertilization. Recently, Baruah and others (2010) have reported a positive correlation between CH₄ emission and photosynthetic activity.



As shown in Figure 4b, an initial emission of N₂O could be detected in all treatments. This peak of N₂O occurred immediately after flushing at a time when the contents of soil NO3- had decreased (Figure 4c). Under these soil redox conditions, both nitrifying and denitrifying organisms could be the main producers of N₂O (Müller and others, 2004). Although there were no significant differences, treatments that had winter cover showed a tendency to reduce their N2O emission before the flood was established compared to soils without cover. In fact, the use of non-legume crops in winter has been described as an effective practice to reduce N2O emissions (Gomes and others, 2009), due to competition with soil microorganisms for the available NO₃. There was no significant effect of N fertilization on N₂O fluxes in any treatment. It is generally accepted that the emission of N2O increases immediately after fertilization in dry soils (Bronson and others, 1997; Yagi and others, 1996; Cai and others, 1997). In the case of rice that was sown directly on the fallow of ryegrass, microorganisms could have immobilized N due to the high C/N ratio of the ryegrass. However, Dobermann and Cassman (2002) suggested that the main factor affecting the emission of N₂O is the N turnover rate, taking into account the synchronization between N mineralization and plant absorption. Fractional application of N, a recommended method of application for this crop, is likely to increase the plant's efficiency of N use (Irisarri and others, 2007), which has an inverse relationship with the N₂O emission (Kroeze and Mosier, 2000). An event of a small flux of N2O emission was measured after draining the field at the end of the crop. This flux could be due to the release of N₂O trapped in the soil and optimal redox conditions for the production of N2O. Non-legume crops have been reported as efficient consumers of residual NO3- in the soil (Gomes and others, 2009) and therefore able to reduce their losses. Thus, when rice cultivation is not occupying the soil, emissions must be measured in order to consider the entire system.

Finally, both experiments, greenhouse and field, showed that the emission of CH_4 coincided with the reproductive stage of rice, while the emission of N_2O was more influenced by agricultural practices such as water management, nitrogen fertilization and previous land use. Although our data were obtained during a single harvest, with particular climatic conditions, the results of the greenhouse and field experiments are consistent.



Seasonal fluxes

Table 4 shows the cumulative fluxes of CH_4 and N_2O throughout the rice crop cycle. While sampling dates are scarce to draw definitive conclusions, the integrated fluxes of CH_4 were in all cases at least 18 times greater than those of N_2O in CO_2 equivalents (in 100 years). Although N_2O is a much more potent greenhouse gas than CH_4 in terms of global warming, its seasonal emission per hectare was much lower.

The Esif (emitted seasonal integrated flux) of CH₄ of fertilized rice after winter covering with ryegrass, was significantly higher than that of the other treatments (Table 4). One possible explanation for this result is that the combination of no-till, ryegrass cover, and nitrogen fertilization may have increased carbon supply to methanogenic organisms (Wassmann and others, 2000). In the case of N₂O, none of the treatments recorded a different seasonal emission than the rest, although the high variability of the Esif may have hidden the potential effects of the treatments. It should also be considered that emission rates may be overestimated since the fluxes were measured in the hottest period of the day (13:00 -15:00 h), when the maximum emission rates occur (Hou and others, 2000).

The median seasonal emissions of other irrigated rice paddies in different parts of the world range from 34 g CH₄ m⁻² (China) to 25 g CH₄ m⁻² (USA). Our seasonal data range from 17 to 21 g CH₄ m⁻² for rice sown on bare soil in winter, and between 32 and 64 g CH₄ m⁻² for the crop sown on ryegrass cover. According to our results, the establishment of a winter ryegrass cover increased the CH₄ flux.

A recent review of emissions of N₂O from various rice paddies reports seasonal averages of 0.667 \pm 0.885 kg N ha⁻¹, revealing the great variability in the flux of this GHG and the consequent difficulty in comparing data (Akiyama and others, 2005). On the other hand, when comparing the emissions per hectare we must consider the high rice yields in Uruguay (8000 - 8500 kg ha⁻¹; ACA 2011) and the average yields of Asian countries (5000 kg ha⁻¹, AFSIS). This would result in lower emissions in CO₂ equivalents per kg of rice in the case of Uruguay, although it would be necessary to increase the sampling dates to validate this conclusion.

Measurements of these gases during the course of the day and in winter would allow obtaining annual emission data that would be comparable to the emissions reported by other countries.

Treatment ²	Fie CH4 (kg C–CH4 ha ⁻¹)	Fie CH4 (CO2 equiv. 100 years)	Fie N2O (kg N–N2O ha ⁻¹) (O	Fie N2O CO2 equiv. 100 years)
Without vegetation	156 ^b	4368	0,5 ª	243
Without vegetation + N	129 ^b	3612	0,4 ª	195
Ryegrass	242 ^b	6776	0,4 ª	195
Ryegrass + N	482ª	13496	1,1 ^a	535

Table 4. Emitted seasonal integrated flux (Esif) of CH₄ and N₂O by rice cultivation in field experiment¹.

¹Values followed by different letters were significantly different (p<0.01).

²Treatment: combination of prior winter cover and nitrogen fertilization (+ N) during rice cultivation.

Conclusions

This first approximation to GHG emission in Uruguayan rice paddies confirmed that CH_4 is the main gas emitted and that the emission patterns of both gases have an opposite behavior throughout the crop cycle. Rice sown on a cover of ryegrass and fertilized with nitrogen emitted more than twice as much CH_4 as rice sown on bare soil. In both the greenhouse and field experiments, the highest emissions of CH_4 coincided with the reproductive stage of rice, while the N₂O emissions peaked at the vegetative phase and were influenced by water management and nitrogen fertilization. These preliminary results on the effect of some crop management practices on GHG emissions reinforce the need for local data, to contribute to the development of the national GHG inventory, the calculation of the C footprint and the design of emission mitigation strategies.

Acknowledgements

This project was funded by INIA-FPTA No. 238. We thank Estefanía Geymonant, Mariana Urraburu, Leticia Pérez and Germán Pérez, for their collaboration in the measurements.



References

ACA. 2011. Datos estadísticos [En línea]. Consultado 12 diciembre 2011. Disponible en:http://www.aca.com.uy/index.php?option=com_ content&view=section&layout=blog&id=3&Itemid=9

Akiyama H, Yagi K, Yan X. 2005. Direct N₂O emissions from rice paddy fields: summary of available data. *Global Biogeochemical Cycles,* 19(1): GB1005.

Baggs EM, Philippot L. 2011. Nitrous oxide production in the terrestrial environment. En: Moir JWB. [Ed.]. Nitrogen cycling in bacteria. Norkfolk: Caister Academic Press. pp. 211-232.

Baggs EM, Smales CL, Bateman EJ. 2010. Changing pH shifts the microbial source as well as the magnitude of N₂O emissions form soil. *Biology and Fertility of Soils* 46: 793 - 805.

Baruah KK, Gogoi B, Gogoi P. 2010. Plant physiological and soil characteristics associated with methane and nitrous oxide emission from rice paddy. *Physiology and Molecular Biology of Plants,* 16: 79 - 91.

Bodelier PL, Rslev P, Henckel T, Frenzel P. 2000. Stimulation by ammonium-based fertilizers of methane oxidation in soils around rice roots. *Nature*, 403: 421 - 424.

Bronson KF, Neue HU, Singh U, Abao EB. 1997. Automated chamber measurements of methane and nitrous oxide flux in a flooded rice soil : I. Residue, nitrogen, and water management. *Soil Society of American Journal*, 61: 981- 987.

Cai Z, Laughlin R, Stevens RJ. 2001. Nitrous oxide and dinitrogen emissions from soils under different water regimes and straw amendment. *Chemosphere,* 42: 113 - 121.

Cai ZC, Xing GX, Yan XY, Xu H, Tsuruta H, Yagi K, Minami K. 1997. Methane and nitrous oxide emissions from rice paddy fields as affected by nitrogen fertilisers and water management. *Plant and Soil*, 196: 7 -14.

Chidthaisong A, Watanabe I. 1997. Methane formation and emission from flooded rice soil incorporated with ¹³C-labeled rice straw. *Soil Biology and Biochemistry*, 29: 1173 - 1181.

Crutzen PJ, Lelieveld J. 2001. Human impact on atmosphere chemistry. *Annual Review of Earth and Planetary Sciences*, 29: 17 - 45.

Deambrosi E. 2003. Rice production system in Uruguay and its sustainability. En: Proceedings of the III International Conference of Temperate Rice; Punta del Este, Uruguay. Montevideo: INIA. p. 19.

Dobermann A, Cassman KG. 2002. Plant nutrient management for enhanced productivity in intensive grain production systems of the United States and Asia. *Plant and Soil*, 247: 153 - 175.

Gomes J, Bayer C, de Souza F, Costa M, de Cássia Piccol Zanatta JA, Beber Vieira FC, Six J. 2009. Soil nitrous oxide emissions in long-term cover crops-based rotations under subtropical climate. *Soil Tillage Research*, 106: 36 - 44.

Holzapfel-Pschorn A, Conrad R, Seiler W. 1986. Effect of vegetation on the emission of methane from submerged paddy soil. *Plant and Soil*, 92: 223 -233.

Hou AX, Chen GX, Wang ZP, Van Cleemput O, Patrick WH. 2000. Methane and nitrous oxide emissions from a rice field in relation to soil redox and microbiological processes. *Soil Science Society of America Journal*, 64: 2180 - 2186.

IPCC. 2007. Climate change 2007 : The physical science basis. Cambridge : Cambridge University Press. pp. 137 - 153.

Irisarri P, Gonnet S, Deambrosi E, Monza J. 2007. Cyanobacterial inoculation and nitrogen fertilization in rice. *World Journal of Microbiology and Biotechnology*, 23: 237 - 243.

Itoh M, Sudo S, Mori S, Saito H, Yoshida T, Shiratori Y, Suga S, Yoshikawa N, Suzue Y, Mizukami H, Mochida T, Yagi K. 2011. Mitigation of methane emissions from paddy fields by prolonging midseason drainage. *Agriculture, Ecosystems and Environment*, 141: 359 - 372.

Jacobson MZ. 2005. Atmospheric pollution: history, science and regulation. New York: Cambridge University Press. pp. 3 - 226.

Kerdchoechuen O. 2005. Methane emission in four rice varieties as related to sugars and organic acids of roots and root exudates and biomass yield. *Agriculture, Ecosystems and Environment,* 108: 155 -163.

Kravchenko IK, Yu K. 2006. Relationship between major soil properties and culturable microorganisms affecting methane and nitrous oxide dynamics in rice soils. *Archives of Agronomy and Soil Science*, 52: 607-615.



Kroeze C, Mosier AR. 2000. New estimates for emission of nitrous oxide. En: van Ham JEA [Ed.]. Non-CO2 Greenhouse gases: scientific understanding, control and implementation. Netherlands : Kluwer Academic Publishers. pp. 45 - 64.

Lindau CW, Bollich PK, Delaune RD, Patrick WH, Law VJ. 1991. Effect of urea fertilizer and environmental factors on CH4 emissions from a Louisiana, USA rice field. *Plant and Soil*, 136: 195 - 203.

Macalady JL, MacMillan AMS, Dickens AF, Tyler SC, Scow KM. 2002.Populations dynamics of type I and II methanotrophs bacteria in rice soils. *Environmental Microbiology*, 4: 148 - 157.

Méndez R, Deambrosi E, Blanco P, Saldain N, Pérez de Vida F, Gaggero M. 2003. Technology for rice seeding with reduced or no-till for the Eastern zone of Uruguay. En: Proceedings of the III International Conference of Temperate Rice; Punta del Este, Uruguay. Montevideo: INIA. p. 64.

Müller C, Stevens RJ, Laughlin RJ, Jäger HJ. 2004. Microbial processes 1 and the site of N2O production in a temperate grassland soil. *Soil Biology and Biochemistry*, 36: 453 - 461.

Mulvaney RL. 1996. Chemical Methods Nitrogen inorganic forms. En: Methods of Soil Analysis : Part 3. Madison : Soil Science Society of America. pp. 1162-1171.

MVOTMA, DINAMA, Unidad de Cambio

Climático. 2010. Tercera comunicación nacional a la conferencia de las partes en la convención marco De las naciones unidas sobre cambio climático: Resumen ejecutivo. Montevideo: MVOTMA. 34 p.

Neue H-U, Sass RL. 1994. Trace gas emissions from rice fields. En: Prinn RG. [Ed.]. Global atmospheric-biospheric chemistry. New York : Plenum Press. pp. 119-147.

Neue H-U, Wassmann R, Kludze HK, Bujun W, Lantin RS. 1997. Factors and processes controlling methane emissions from rice fields. *Nutrient Cycling in Agroecosystems*, 49: 111-117.

Perdomo C, Irisarri P, Ernst O. 2009. Nitrous oxide emissions from an uruguayan argiudoll under different tillage and rotation treatments. *Nutrient Cycling in Agroecosystems*, 84: 119 - 128.

Schutz H, Holzaptel-Pschorn H, Conrad R, Rennenberg H, Seiler W. 1989. A three-year continuous record on the influence of day time season and fertilizer treatment on methane emission rates from an Italian rice paddy. *Journal of Geophysical Research*, 94: 16405-16416.

Smith KA, Ball T, Conen F, Dobbie KE, Massheder J, Rey A. 2003. Exchange of greenhouse gases between soil and atmosphere : interactions of soil physical factors and biological processes. *European Journal of Soil Science*, 54: 779 - 791.

Terra J, Sanchez A, Deambrosi E, Méndez R. 2009. Efecto de cultivos de cobertura invernal sobre la respuesta a N en el cultivo de arroz en siembra directa. En: Arroz : Resultados experimentales 2009 - 2010. Montevideo : INIA. (Actividades de Difusión ; 611). pp. 9 - 20.

Towprayoon S, Smakgahn K, Poonkaew.S 2005. Mitigation of methane and nitrous oxide emissions from drained irrigated rice fields. *Chemosphere*, 59: 1547 - 1556.

USDA. 1998. Keys to Soil Taxonomy. 8th ed. Washington : USDA. 326p.

Wassmann R, Neue HU, Lantin RS, Makarim K, Chareonsilp N, Buendia LV, Rennenberg H. 2000. Characterization of methane emissions from rice fields in Asia : II. Differences among irrigated, rainfed and deepwater rice. *Nutrient Cycling in Agroecosystems*, 58: 13 - 22.

Wassmann R, Papen H, Rennenberg H. 1993. Methane emission from rice paddies and possible mitigation strategies. *Chemosphere*, 26: 201 - 217.

Watanabe T, Chairoj P, Tsuruta H, Masarngsan W, Wongwiwatchai C, Wonprasaid S, Cholitkul, W, Minami K. 2000. Nitrous oxide emissions from fertilized upland fields in Thailand. *Nutrient Cycling in Agroecosystems*, 58: 55 - 65.

Xing GX, Shi SL, Shen GY. 2002. Nitrous oxide emissions from paddy soil in three rice based cropping systems in China. *Nutrient Cycling in Agroecosystems*, 64: 35 - 43.

Yagi K, Minami K. 1990. Effect of organic matter application 1 on methane emission from some Japanese paddy fields. *Soil Science and Plant Nutrition,* 36: 599 – 610.

Yagi K, Tsuruta H, Kanda K, Minami K. 1996. Effect of water management on methane emission from a Japanese rice field: automated methane monitoring. *Global Biogeochemical Cycles*, 10: 255 – 267.